



Toulouse, January 04<sup>th</sup> 2023

**STUDY 22-3283**

**REPORT 23-1983**

**EVALUATION OF THE VIRUCIDAL ACTIVITY OF ON PLASTICS AND OTHER  
NON-POROUS SURFACES AGAINST HUMAN CORONAVIRUS 229E ACCORDING  
TO THE METHODOLOGY OF STANDARD ISO 21702 MAY 2019**

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## I- TEST LABORATORY IDENTIFICATION

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## II -SAMPLE IDENTIFICATION

- Name of support: CLEAR DIAMOND ANTI-VIRAL  
- Batch number: CD810AV  
- Date of receipt: December/06/2022  
- Internal code: 22-3283-2

- Name of support: CONTROL PIECES  
- Batch number: CD858S  
- Date of receipt: December/06/2022  
- Internal code: 22-3283-1

- Supplier: Hi-Bond Tapes Ltd

- Period of testing: December 2022

## III - TEST METHOD

### III-1 VIRUS

Name: Human Coronavirus 229E  
Origin: ATCC  
Reference: VR-740  
Supplier batch number: 58505270  
Internal batch number: SS-2-210920 (Passage N°2)

### II-2- Recipient cells

Name: Vero Cells  
Origin: ATCC  
Reference: CCI-81  
Supplier batch number: 3372621  
Internal batch number: WCB-101215 (Passage N°16)

#### **IV -TEST CONDITIONS**

- Contact times: 30 minutes and 1 hour
- Test temperature: 25°C ± 1°C

#### **V- TEST METHOD**

##### **V-1 Control of cytotoxicity**

2.5 ml of neutralizing medium are added to 3 untreated and 3 treated samples. The samples are washed 4 times with the neutralizing medium.

A ten-fold serial dilution is made to check the absence of cellular cytotoxicity.

##### **V-2 Control of the sensitivity of the cells to the virus and stopping the antiviral activity**

2.5 ml of neutralizing medium are added to 3 untreated and 3 treated samples.

The samples are washed 4 times with the neutralizing medium. Then 1.98 ml of recovery medium are mixed with 20 µl of the virus suspension prepared at a concentration of 4 to 6 .10<sup>5</sup> TCID<sub>50</sub>/ml. After 30 min of 25°C incubation, tubes with virus solution are maintained in ice before titration.

##### **V-3 Contact virus/surface**

Each sample with a surface area of 5 cm x 5 cm (control and test samples) is placed in a sterile glass Petri dish.

- 400 µl of the viral suspension are deposited on each surface and spread over 16 cm<sup>2</sup> using a 4 x 4 cm film to reduce desiccation of the inoculum.

##### **V-4 Recovery of the viral film**

After incubation, 3.6 ml of a neutralizing solution (frozen culture medium) are added to the samples in order to recover viable viruses.

The titration of the remaining viable viruses is then carried out immediately.

##### **V-5 Viral titer**

The titration technique is indicated in the standard NF EN 14476 + A2 (July 2019).

A ten-fold serial dilution of the viral suspensions is made in the cell culture medium in neutral glass tubes in order to limit the phenomena of virus adsorption on the surfaces.

Titration is performed on 96-well microplates. Each dilution is transfer in 8 wells.

## **V-6 Viral titer calculation**

The assay is performed by the microplate method of suspension cells. The cytopathic effect is determined at least 4 days of culture.

The number of infectious units is estimated with the SPEARMAN-KÄRBER method by calculating the negative logarithm of the 50% limit point ( $\lg\text{TCID}_{50}$ ) using the following formula:

$\lg\text{TCID}_{50} = \text{Negative logarithm of the highest concentration of virus used} - [(\text{Sum of \% assigned to each dilution}/100 - 0.5) \times (\lg \text{ of dilution})]$

The following tests are carried out 3 times.

## **VI- RESULTS**

### **VI-1 Validation**

#### **VI-1-1 Control of cytotoxicity**

No cytotoxicity was observed on the cells after contact of the culture medium with treated and untreated samples.

#### **VI-1-2 Control of the sensitivity of cells to viruses and cessation of virucidal activity**

The difference between the average titers ( $\lg \text{TCID}_{50}$ ) of the neutralizing solution controls and the sensitivity titers average of the treated and untreated surfaces must be less than or equal to 0.5 lg.

#### **Neutralizing solution control**

- Control 1 :  $\lg \text{TCID}_{50} = 3.88$
- Control 2 :  $\lg \text{TCID}_{50} = 3.75$
- Control 3 :  $\lg \text{TCID}_{50} = 3.75$

**$\lg \text{TCID}_{50}$  neutralizing solution control average = 3.79**

#### **Control Sensitivity of untreated surfaces**

- Control 1 :  $\lg \text{TCID}_{50} = 3.50$
- Control 2 :  $\lg \text{TCID}_{50} = 3.63$
- Control 3 :  $\lg \text{TCID}_{50} = 4.00$

**$\lg \text{TCID}_{50}$  Sensitivity of untreated surfaces average = 3.71**

**Titer neutralizing solution average - Sensitivity of untreated surfaces average = 0.08**

Difference  $\leq 0.5 \lg$  (verification valid)

**Test Sensitivity of treated surfaces**

- Control 1 : lg TCID<sub>50</sub> = 3.88
- Control 2 : lg TCID<sub>50</sub> = 3.38
- Control 3 : lg TCID<sub>50</sub> = 3.63

**lg TCID<sub>50</sub> Control Sensitivity average = 3.63**

**Titer neutralizing solution average - Sensitivity of treated surfaces average = 0.16**

Difference ≤ 0.5 lg (verification valid)

**VI-1-3 TO control**

- Control 1 : lg TCID<sub>50</sub> = 5.50
- Control 2 : lg TCID<sub>50</sub> = 6.00
- Control 3 : lg TCID<sub>50</sub> = 5.38

**lg TCID<sub>50</sub> TO average = 5.63**

Maximum viral title - Minimum viral title = 0.11

Average of the 3 viral titles

The titer (lg DICT<sub>50</sub>) of the 3 tests at TO must be homogeneous

Maximum viral titer - Minimum viral titer / Average of the 3 viral title ≤ 0,2.

**TCID<sub>50</sub> average /ml = 4.27 10<sup>6</sup>**

Average TCID<sub>50</sub> /ml = 10<sup>average log<sub>10</sub> DCICT<sub>50</sub></sup> × 10

**Infectivity titer (TCID<sub>50</sub>/cm<sup>2</sup>) =**

$$\frac{\text{TCID}_{50}/\text{ml} * \text{Volume de récupération (4ml)}}{\text{Surface (16 cm}^2\text{)}} = 1.10^6$$

Infectivity titer at TO (TCID<sub>50</sub>/cm<sup>2</sup>) must be between 8.94 10<sup>5</sup> and 4.46 10<sup>6</sup>

**VI-2 Tests**

**VI-2-1 Control**

- Control 1 : lg TCID<sub>50</sub> = 5.38
- Control 2 : lg TCID<sub>50</sub> = 5.38
- Control 3 : lg TCID<sub>50</sub> = 5.63

**lg TCID<sub>50</sub> TO average = 5.46**

**Moyenne TCID<sub>50</sub> /ml = 2.88 10<sup>6</sup>**

Moyenne TCID<sub>50</sub> /ml = 10<sup>Moyenne log<sub>10</sub> DCICT<sub>50</sub></sup> × 10

**Infectivity titer (TCID<sub>50</sub>/cm<sup>2</sup>) =**

$$\frac{\text{TCID}_{50}/\text{ml} * \text{Recovery volume (4ml)}}{\text{Surface (16 cm}^2\text{)}} = 7.21 \cdot 10^5$$

Infectivity titer at maximum contact time (1 hour) (TCID<sub>50</sub>/cm<sup>2</sup>) must be greater than 2.21 10<sup>3</sup>

#### **VI-2-2 Test 30 minutes**

- Assay 1 : lg TCID<sub>50</sub> = 2.88
- Assay 2 : lg TCID<sub>50</sub> = 2.88
- Assay 3 : lg TCID<sub>50</sub> = 3.13

**lg TCID<sub>50</sub> Assay average = 2.96**

**R = lg TCID<sub>50</sub> control maximum contact time average - lg TCID<sub>50</sub> Test 30 minutes average = 2.50 log**

#### **VI-2-3 Test 1 hour**

- Assay 1 : lg TCID<sub>50</sub> = 2.50
- Assay 2 : lg TCID<sub>50</sub> = 2.63
- Assay 3 : lg TCID<sub>50</sub> = 1.88

**lg TCID<sub>50</sub> Assay average = 2.34**

**R = lg TCID<sub>50</sub> control maximum contact time average - lg TCID<sub>50</sub> Test 1 hour average = 3.12 log**

### **VII-CONCLUSION**

According to the methodology of the ISO 21702 standard (May 2019), contact of treated support CLEAR DIAMOND ANTI-VIRAL batch CD810AV with the strain of Human Coronavirus 229E induces a reduction of the viral titer of 2.50 lg at contact time 30 minutes and 3.12 lg at contact time 1 hour.

The treatment of supports CLEAR DIAMOND ANTI-VIRAL batch CD810AV induces a reduction of the viral load of 99.68% at contact time 30 minutes and 99.92% at contact time 1 hour.